

FORENSIC SCIENCE

# Validation of PowerPlex<sup>®</sup> 18D Kelly Borycki, BS<sup>1</sup>; Jennifer Bas, MFS<sup>2</sup>; Justin Godby, MS<sup>1</sup>; Pamela Staton, PhD<sup>1</sup>

# Abstract

This study sought to validate the performance of the PowerPlex<sup>®</sup> 18D direct amplification kit (PP18D) for use with single-source FTA<sup>®</sup> samples.

PP18D was determined to yield full profiles from samples with concentrations as low as  $0.4ng/\mu L$ . The kit was shown to be precise at all 18 loci. Known samples were collected from LVMPD employees, and all samples were concordant. Seven samples that were amplified and run on different instruments and different days to test reproducibility were also concordant. Other common sample substrates (cotton swabs, Omni<sup>®</sup> swabs, extracted DNA) were also tested and a full profile was obtained from all samples.

### Introduction

The purpose of this study was to ensure that the PowerPlex<sup>®</sup> 18D kit, when used for direct amplification of FTA<sup>®</sup> card samples, would produce DNA profiles of acceptable quality to be entered into CODIS.

This study will impact the forensic science community by showing the reliability of a new technology that can potentially save a significant amount of time and money during the production of forensic DNA profiles. It will also provide other laboratories considering adopting direct amplification with a general internal validation scheme.

### **Materials and Methods**

• Samples collected using Whatman<sup>®</sup> EasiCollect<sup>TM</sup> devices

• Punches of the FTA<sup>®</sup> cards taken using the Harris .2mm Manual Punch

• Half reaction amplification  $(7.5\mu L \text{ water}, 2.5\mu L)$ 5X Master Mix, 2.5µL 5X Primer Pair Mix per sample)

•27 cycle amplification on Applied Biosystems GeneAmp<sup>®</sup> PCR System 9700 thermal cyclers

• Capillary electrophoresis on Applied Biosystems 3130x1 Genetic Analyzers

• Applied Biosystems GeneMapper<sup>®</sup> ID-X v1.1.1 used for data analysis

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Sensitivity and Stochastic				
Calvin	Rio	Luxor	• 3kV/5sec for Calvin	
5 ng	5 ng	5 ng		
4 ng	4 ng	4 ng	• 3kV/3sec for Rio and	
3 ng	3 ng	3 ng	Luxor	
2 ng	2 ng	2 ng	* Due to instrument maintenance, a stochastic threshold is currently being established by LVMPD	
1 ng	1 ng	1 ng		
0.8 ng	0.8 ng	0.8 ng	Offscale	
0.6 ng	0.6 ng	0.6 ng	peaks	
0.4 ng	0.4 ng	0.4 ng	Good profile Dropout	
0.2 ng	0.2 ng	0.2 ng		
0.1 ng	0.1 ng	0.1 ng		
Relative sensitivity of three 3130xl genetic analyzers				

# **Database-Type Samples**

• Extracts from cotton-tipped swabs using DNA IQ<sup>TM</sup> with Biomek<sup>®</sup> NX

> Average quant value (0.152ng/µL) x Volume added to amp (7.5µL) Average target (1.14ng)

• Full profiles below 5ng recommended input

# **Known/Non-Probative** and Reproducibility

- 65 samples compared to LVMPD staff database
- 7 samples processed on
  - 3 separate days
  - Different thermal cyclers
  - Different genetic analyzers

### Concordance

• All profiles obtained in this validation matched a known profile from the same donor

• NIST samples yielded expected profiles

• No null alleles or primer binding site mutations detected





# **Casework-Type Samples**

- Manual extraction of buccal swabs
  - PrepFiler<sup>TM</sup>
  - Organic
  - Qiagen
  - Chelex<sup>®</sup>
- 5ng amplification target
- Full profiles

Sample Type

• Full profile from manual FTA<sup>®</sup> cards at 27 cycles

•Partial profile from cotton and Omni<sup>®</sup> swabs at 27 cycles

• Full profile from cotton and Omni<sup>®</sup> swabs at 28 cycles without lysis buffer



http://www.evidentcrimescene.com/cata/blood/swabster2.jpg



http://www.fishersci.com/ecomm/servlet/fsproductdetail 10652 1631910 29104 -1 0

# Contamination

• All negative controls and reagent blanks contained no evidence of contamination

• One mixed sample – minor peaks found to match neighboring sample; attributed to human error

- available.

- validation.

# Conclusions

• PowerPlex<sup>®</sup> 18D is a robust kit that consistently produced accurate and reproducible results.

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• PowerPlex<sup>®</sup> 18D will be adopted by the LVMPD Biology/DNA Detail for use with database samples.

• Further studies will be done on non-FTA<sup>®</sup> card samples once the lysis buffer becomes commercially

### References

Oostdik, K., Ensenberger, M., Krenke, B., Spreecher, C., and Storts, D. The PowerPlex® 18D System: A Direct Amplification STR System with Reduced Thermal Cycling Time. Profiles in DNA 2011; 14(1) [Internet].

French, J. Implementation of the PowerPlex® 18D System in a Databasing Laboratory: Transforming Operations to Improve Efficiency. Profiles in DNA 2011; [Internet]

Wieczorek, D. and Krenke, B. Direct Amplification from Buccal and Blood Samples Preserved on Cards Using the PowerPlex® 16 HS System. Profiles in DNA 2009; [Internet].

Suzanne, E., Julien, S., Hernandez, M., Menager, C., and Pene, L. Direct Amplification from Saliva and Blood Samples Preserved on Cards Using the PowerPlex® ESX 16 System. Profiles in DNA 2010; [Internet].

Butler, J. Forensic DNA Typing: Biology, Technology, and Genetics of STR Markers. Second edition. London: Elsevier, 2005.

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