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Development of an Identification and Derivatization Method for Synthetic Cathinones by GC-MS using Perfluoroacyl Anhydrides

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Abstract

Synthetic cathinones have become increasingly popular in the past decade. Three synthetic cathinones, Mephedrone, Methylone, and MDPV have been placed into Schedule I of the Controlled Substances Act. The Analog Act was created to allow substances to be scheduled if they were similar to an already scheduled compound. Synthetic cathinones have the potential for positional isomers, which produce ambiguous mass spectra. Derivatization has been proven useful for determining differences in the mass spectra of similar compounds. The effect of three perfluoroacyl derivatizing agents on synthetic cathinone standards was tested along with the ability to differentiate positional isomers within a mixture. Compounds containing a primary or secondary nitrogen readily derivatized; the compounds containing a tertiary nitrogen, however, were not able to be derivatized. When placed into mixtures, the positional isomers were distinguished nearly every time. Future studies include the determination of a method to derivatize synthetic cathinones containing a tertiary nitrogen.

Introduction

Synthetic cathinones, more commonly referred to as bath salts, are becoming increasingly abused in the United States. As a result, forensic drug laboratories are receiving an increasing number of samples believed to contain these compounds. Bath salts are gaining popularity due to their psychoactive and stimulant properties, which are similar to those of amphetamine and cocaine. Because these are synthetic compounds, their molecular structure can be slightly altered to circumvent scheduling under the United States Drug Enforcement

Administration's (DEA's) Controlled Substances Act. In most states, and federally, a drug is scheduled based on its exact structure; therefore, an alteration of a single position in the molecule could yield a new, legal drug that would theoretically have the same effects as the original scheduled drug. However, the Federal Analog Act of 1986 states that any compound that is "substantially similar" to a Schedule I or II controlled substance, has similar pharmacological effects, and is intended for human consumption is to be treated as if it were also scheduled (1). Most of the packaging for bath salts contains the disclaimer "not for human consumption" in an effort to avoid the Federal Analog Act. Most states require that only two conditions of the Analog Act are met to identify the substance as an analog. In October 2011, the DEA placed an emergency ban on Methylone and the Synthetic Drug Abuse Prevention Act of 2012 (S.3187 Subtitle D) of the Food, Drug, and Cosmetic Act added two new methcathinones to Schedule I, 4-Methylmethcathinone (mephedrone) and 3,4-Methylenedioxypyrovalerone (MDPV) (2).

Due to the minor structural variations and legal ramifications associated with synthetic cathinones, identification of the exact structure is imperative. Not only must the presence of a cathinone be documented, but the exact compound must be identified to know if it is scheduled, which often means differentiating between positional isomers. Gas chromatography-mass spectrometry (GC-MS) is the primary method used in a forensic laboratory to confirm the identity of a drug. Generally, cathinones differ due to varying functional groups on the benzene ring. When the underivatized compound is fragmented in the electron source of the MS, the benzene ring is neutralized, and therefore not recognized by the detector, yielding mass spectra with the same ions in the same relative ratios.

Perfluoro acyl anhydrides have been used in the past to differentiate cathinones and similar compounds, like the amphetamines, by GC-MS (3,4). The addition of an acyl group to an amine lowers the basicity of the compound, favoring alternative fragmentation pathways to produce more diagnostic mass spectra (5). Further, perfluoro acyl anhydrides help to

withdraw electron groups from the benzene ring, making it less stable and a better candidate for ionization. Trifluoroacetic anhydride (TFAA), heptafluorobutyric anhydride (HFBA), and pentafluoropropionic anhydride (PFPA) are common perfluoroacyl derivatizing agents and were used in this research to derivatize 23 synthetic cathinones. The perfluoro acyl anhydrides bond with nitrogen atoms contained in each of the synthetic cathinones. Some contain a pyrrolidine ring which makes the nitrogen tertiary, lacking the hydrogen necessary for the reaction with the perfluoroacyl anhydride (6). Pyridine was used in an attempt to open the ring allowing the anhydride to bond with the nitrogen. The effects of the derivatizing agents on each compound and several mixtures of multiple cathinones were compared to determine which agent performed the best for each compound and the best overall by examining the GC retention times, the amount of breakdown products present, and the mass spectra. If the derivatizing agents are able to bond to the nitrogen of the synthetic cathinones, then the mass spectra of the cathinones will be differentiated.

Materials and Methods

Reagents, Standards, Equipment, and Instrumentation

Methanol was purchased from Fisher Scientific (Pittsburgh, PA). Ethyl Acetate and Pyridine were purchased from Sigma-Aldrich (St Louis, MO). TFAA, HFBA, and PFPA were purchased from Fluka Analytical, which is now owned by Sigma-Aldrich. Hydrochloride standards of Butylone, Methedrone, 2-Fluoromethcathinone, 3-Fluoromethcathinone, 4-Fluoromethcathinone, Pentedrone, 2-Methylmethcathinone, 3-Methylmethcathinone, 3,4-Dimethylmethcathinone, 2,3-Methylenedioxymethcathinone, 2-Methoxymethcathinone, 3-Methoxymethcathinone, α -Pyrrolidinopropiophenone, 2-Methylpyrrolidinopropiophenone, 3-Methylpyrrolidinopropiophenone, 4-Methylpyrrolidinopropiophenone, 4-Methoxypyrrolidinopropiophenone, and 3,4-Methylenedioxypyrrolidinopropiophenone, 4-Methoxypyrrolidinopropiophenone, Methylenedioxypyrrolidinopropiophenone, 4-

purchased from Cayman Chemical (Ann Arbor, MI). The gas chromatograph/mass spectrometer with quadrupole ion trap was purchased from Agilent Technologies (Santa Clara, CA). The column is a Zebron ZB-DRUG-1 column from Phenomenex (Torrance, CA). *Sample Preparation*

Each synthetic cathinone standard was analyzed by GC-MS in four different ways: as a standard in methanol, derivatized with TFAA, with HFBA, and with PFPA. The structures for each standard and derivatizing agent are shown in Figs. 1 and 2, respectively. Mixtures were also prepared and analyzed by GC-MS. The standard solutions for analysis were all prepared as follows. Using a Pasteur pipette, approximately 1.0 mg (ranging from 0.4 mg to 2.2 mg) was taken from the Cayman Chemical vial and added to a 350 μ L vial insert inside of a labeled GC vial. Methanol was then flushed down the pipette to ensure the entire standard was inside the insert, which was then filled with methanol and the vial capped.



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Fig. 2: Structures of TFAA, HFBA, PFPA, and Pyridine.

Derivatizations were performed with the same method for all three derivatizing agents. Work was performed under a hood because the reagents have a strong smell and can be harmful if inhaled. The derivatization procedure was adapted from that used by Abdel-Hay et al (5) and Lum (7). A Pasteur pipette was used to sample approximately 1.0 mg of standard. The pipette was placed in a labeled GC vial and 25 μ L of ethyl acetate was used to flush the sample into the vial and serve as the solvent. TFAA, HFBA, or PFPA at a volume of 100 μ L was also added to the vial. The vial was capped and allowed to react for 15 minutes at room temperature. During this time, the vial was placed on top of the GC to add a small amount of heat to aid the derivatization process. The sample was then transferred to a larger, labeled vial under the hood for evaporation. The sample vial was placed on a hot plate on low to facilitate the evaporation process. The resulting residue was reconstituted with 300 μ L of ethyl acetate and the resulting solution transferred into an insert in a labeled GC vial and capped.

Eight of the twenty-three standards were tertiary amines containing a pyrrolidine ring. Derivatization of tertiary amines is not possible using perfluoroacyl anhydrides. Two methods were attempted to derivatize these compounds. Pyridine was used in this process to possibly enolize the carbonyl and create a point of attachment for the perfluoroacyl anhydrides. In the first method, approximately 1.0 mg of cathinone standard was added to a GC vial using a

Pasteur pipette and 25 μ L of ethyl acetate as described previously. Pyridine (300 μ L) was then added to the mixture in the GC vial, followed by 100 μ L of TFAA. The solution turned a dark orange and smoked due to the addition of the acid. The vial was capped and incubated at room temperature for 15 minutes. The liquid was then transferred to a larger, open vial and evaporated under the hood. After a small amount of solvent evaporated, an orange gel had formed. Two solvent washes of 300 μ L each of ethyl acetate were unsuccessfully used to eliminate the gel. This sample was not analyzed on the GC-MS. The second method tested was a slight variation on the first method. After the addition of 300 μ L of pyridine, the vial was capped and incubated at room temperature for 15 minutes and then transferred to a larger vial for evaporation. The residue was reconstituted with 50 μ L of ethyl acetate and the resulting solution was transferred to a new GC vial. TFAA or HFBA (derivatization with PFPA was not attempted) were added to the vial at a volume of 100 μ L. The vial was capped and incubated at room temperature for 15 minutes. The reaction mixture was transferred to a larger vial for evaporation to dryness. The remaining residue was reconstituted with 300 μ L ethyl acetate and the resulting solution transferred to a vial insert in a labeled GC vial and capped.

Mixture samples containing positional isomers were prepared by combining standards previously analyzed to a final volume of 300 µL depending on the number of possible positional isomers. Four mixture groupings were prepared: 2-, 3-, and 4-Fluoromethcathinone; 2- and 3-Methoxymethcathinone and Methedrone; 2- and 3-Methylmethcathinone and Mephedrone; and 2,3-Methylenedioxymethcathinone and Methylone. Compounds that contain a methylenedioxy substitution have only two positional isomers. Each mixture was prepared in quadruple to be analyzed as a standard mixture and TFAA, HFBA, and PFPA derivative mixtures.

Sample Analysis

All samples were analyzed on an Agilent Technologies 6890N Network GC System coupled to a 5973Network Mass Selective Detector equipped with a 7683 Series Injector

autosampler. Separation was achieved using a ZB-DRUG-1 capillary column

(10m×0.18mm×0.18µm). The method parameters are a combination of those used at KSP

and parameters in a study by Davies, Ramsey, and Archer (2009).

Carrier Gas	Helium			
Flow Rate	0.6 mL/min			
Split Ratio	20:1; 35:1; 50:1*			
Injector Temperature	250 °C			
Transfer Line Temperature	290 °C			
Solvent Delay	0.42 min			
Ionization Voltage	0 eV			
Mass Range	40 - 550 m/z			

Table 1: GC-MS Parameters

*The split ratio was chosen according to the concentration of each sample.

Table 2: Oven Temperature Program

Rate	Starting Temperature	Final Temperature	Hold Time	Final Time
~	140 °C	140 °C	6.1 min	6.1 min
20 °C/min	140 °C	200 °C	0.0 min	9.1 min
40 °C/min	200 °C	300 °C	14.1 min	25.7 min

Results & Discussion

Fluoromethcathinones

The total ion chromatograms (TICs) of 2-Fluoromethcathinone (2-FMC), 3-

Fluoromethcathinone (3-FMC), 4-Fluoromethcathinone (4-FMC), and the

Fluoromethcathinone mixture (Fluoro Standard Mix) are shown in Fig. 3. All mass spectral results were library searched using the three libraries available at KSP: SWGDRUG, Cayman Chemical, and Cerilliant. The TIC for 2-FMC contains five labeled peaks (Fig. 3A). 2-FMC is known to decompose during GC-MS analysis (8). The peaks at 0.940 and 1.041 minutes are precursors of 2-FMC (the mass spectra of which can be seen in Appendix A Figs. A1 and A2, respectively) that were matched with data in the SWGDRUG Monograph (8). The peak at 1.422 minutes is 2-FMC, while the peak at 1.590 minutes is the 2-FMC enamine (mass spectrum in Fig. A3). An enamine occurs when there is ring closure between the benzene ring substitution and the methyl group on the side chain, resulting in the loss of two hydrogens.

The base peak for the enamine is two m/z units lower than the base peak of the synthetic cathinone. Using the base peak value allows for identification of the enamine. Lastly, the peak at 3.94 minutes is the result of ring closure after the loss of HF, the mass spectrum of which can be found in Fig. A4, also consistent with SWGDRUG results. The TIC for 3-FMC also has five peaks (Fig. 3B). An expanded view of the chromatogram for 3-FMC can be seen in Fig. A5. The peaks at 0.825 and 1.177 minutes are precursors of 3-FMC (the mass spectra of which can be seen in Figs. A6 and A7, respectively). The mass spectra of these are consistent with the mass spectra of the 2-FMC precursors. The mass spectrum of the peak at 1.577 minutes is breakdown of 3-FMC (mass spectrum in Fig. A8). The peak at 1.518 minutes is 3-FMC and the peak at 1.636 minutes is the 3-FMC enamine (Fig. A9). The chromatogram for 4-FMC has only one peak, which is the actual 4-FMC (Fig. 3C). Fig. 3D shows the chromatogram of the mixture of the three Fluoromethcathinone standards. Appendix A shows an expanded view of the chromatogram (Fig. A10). There are several peaks in this mass spectrum. The peaks at 1.000, 1.791, 3.471 minutes, and all the peaks afterward are all column bleed and due to low sample concentration you can see this column bleed. The peaks at 0.830, 0.935, 1.039, and 1.178 minutes are all breakdown or rearrangements that were seen when the standards were run on their own. 2-FMC elutes at 1.423 minutes, 3-FMC elutes at 1.499 minutes, and 4-FMC elutes at 1.572 minutes (mass spectra in Figs. A11-A13, respectively). The similarity between the mass spectra of the Fluoromethcathinone standards can be observed in Fig. 4. Table 3 summarizes the data for the Fluoromethcathinone isomers including the base peak for each mass spectrum and the ion ratios of the base peak abundance to the abundance of the major ions. 2-FMC shows the most difference with the change in ratio of the 75, 95, and 123 m/z ions compared that of the 3-FMC and 4-FMC mass spectra. 3-FMC and 4-FMC show little difference. Comparing between these mass spectra, it would be difficult to match an unknown compound present in a forensic sample to a known standard.





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Table 3: Flu	oromethcathinone I	Data
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Retention	Base Peak	Major Ions in m/z (Base Peak
Time	(Abundance)	Abundance/Ion Abundance Ratio)
1.42 min	58 m/z	42(20), 69(63), 75(12), 95(10), 123(11),
	(305856)	161(55), 181(495)
1.51 min	58 m/z	42(23), 69(60), 75(11), 95(6.7), 123(15),
	(1217932)	166(71), 181(1030)
1.56 min	58 m/z	42(23), 69(63), 75(11), 95(6.7), 123(11),
	(227685)	166(101), 181(135)
2.75 min	154 m/z	42(9.1), 56(23), 69(264), 95(8.1),
	(950464)	110(3.8), 123(2.4), 155(17), 164(51),
		277(381)
2.31 min	154 m/z	42(10), 56(26), 69(13), 95(6.4), 110(3.8),
	(1491968)	123(4.3), 155(17), 164(55), 277(73)
2.27 min	154 m/z	42(8.9), 56(23), 69(11), 95(4.3),
	(169095)	110(3.7), 123(1.3), 155(16), 164(44),
	271	277(41)
$2.49 \mathrm{mm}$	254 m/z	42(13), 56(24), 69(20), 95(12), 109(110),
	(1324032)	123(4.1), 169(16), 210(3.9), 255(12),
0.10	074 /	377(2085)
$2.19 \mathrm{min}$	254 m/z	42(15), 56(27), 69(21), 95(6.0), 109(83),
	(2651648)	123(4.2), 169(9.7), 210(2.2), 255(7.8),
0.00 min	054 m /=	377(136)
2.09 min	254 m/z	42(12), 56(23), 69(17), 95(6.9), 109(86),
	(391360)	125(2.2), 169(16), 210(4.0), 255(12), 377(154)
2 20 min	204 m/z	42(11) = 6(22) + 69(41) + 75(20) + 95(10)
2.35 mm	(1313792)	(42(11), 56(25), 65(41), 75(20), 55(10), 109(135), 123(34), 160(36), 205(15)
	(1010102)	327(838)
2 07 min	204 m/z	42(11) 56(24) 69(40) 75(20) 95(77)
2.07 11111	(899732)	109(158) $123(54)$ $160(35)$ $205(14)$
	(000102)	327(139)
2.00 min	204 m/z	42(11), 56(25), 69(38), 75(17), 95(5.8)
	(1170944)	109(127), 123(1.8), 160(3.7), 205(15).
	()	327(81)
	Retention Time 1.42 min 1.51 min 1.56 min 2.75 min 2.31 min 2.27 min 2.49 min 2.19 min 2.39 min 2.30 min 2.00 min	Retention TimeBase Peak (Abundance)1.42 min $58 m/z$ (305856)1.51 min $58 m/z$ (1217932)1.56 min $58 m/z$ (227685)2.75 min $154 m/z$ (950464)2.31 min $154 m/z$

Fig. 5 displays the TICs for the TFAA derivatives of the Fluoromethcathinones, each of which contains one peak that is the derivative itself. The retention times for each TFAA derivatized Fluoromethcathinone along with the major ions and their ratios are given in Table 3. The TIC in Fig. 5D displays the TFAA derivative mixture (Fluoro TFAA Mix) of the Fluoromethcathinones. 3-FMC-TFAA and 4-FMC-TFAA co-eluted at 2.316 minutes as demonstrated by the mass spectrum in Fig. A14. The co-elution is due to the similarity of the retention times between the two compounds. The mass spectrum for 2-FMC-TFAA, which

elutes at 2.750 minutes in the mixture, is in Fig. A15. The mass spectra of the TFAA derivatives of the Fluoromethcathinones are shown in Fig. 6. The major ions present in all three mass spectra were the same, but in different relative abundances, which allowed for differentiation.



Fig. 5: Total Ion Chromatograms of TFAA Derivatives of the Fluoromethcathinones. A: 2-FMC-TFAA; B: 3-FMC-TFAA; C: 4-FMC-TFAA; D: Fluoro TFAA Mix.



Fig. 6: Mass spectra of TFAA Derivatives of the Fluoromethcathinones. A: 2-FMC-TFAA; B: 3-FMC-TFAA; C: 4-FMC-TFAA.

The TICs for the HFBA derivatives of the Fluoromethcathinones are presented in Fig. 7. 2-FMC-HFBA elutes at 2.498 minutes (Fig. 7A). The chromatogram in Fig. 7B contains three peaks. The peak at 1.498 minutes is a small amount of underivatized 3-FMC (mass spectrum in Fig. A16). The peaks at 2.099 and 2.197 are both 3-FMC-HFBA. The concentration of 3-FMC-HFBA is high in this sample and this can lead to peak fronting. 4-FMC-HFBA elutes at 2.096 minutes (Fig. 7C). The TIC of the Fluoromethcathinone HFBA mixture (Fluoro HFBA Mix) can be seen in Fig. 7D; an expanded chromatogram can be seen in Fig. A17. A small amount of underivatized 3-FMC can be seen at 1.498 minutes in the mixture. 4-FMC-HFBA elutes at 2.095 minutes, 3-FMC-HFBA elutes at 2.177 minutes, and 2-FMC-HFBA elutes at 2.484 minutes (mass spectra in Figs. A18-A20, respectively). As with the TFAA derivatives, mass spectra of the HFBA derivatives (Fig. 8) all have the same major ions, but in different relative ratios; making them easier to distinguish from one another in comparison to the underivatized mass spectra. The ions and their ion ratios compared to the base peak are given in Table 3.



Fig. 7: Total Ion Chromatograms of HFBA Derivatives of the Fluoromethcathinones. A: 2-FMC-HFBA; B: 3-FMC-HFBA; C: 4-FMC-HFBA; D: Fluoro HFBA Mix.



FMC-HFBA; C: 4-FMC-HFBA.

Fig. 9 shows the total ion chromatograms for the PFPA derivatives of the Fluoromethcathinones. 2-FMC-PFPA (Fig. 9A) has two peaks. The peak at 2.393 minutes is 2-FMC-PFPA and the peak at 11.179 minutes is either air or vial cap bleed due to the low molecular ion and simple mass spectrum. 3-FMC-PFPA (Fig. 9B) contains two peaks. The peak at 1.496 minutes is a small amount of underivatized 3-FMC (mass spectrum in Fig. A21) and the peak at 2.073 is 3-FMC-PFPA. 4-FMC-PFPA (Fig. 9C) includes one peak at 2.004 minutes, the PFPA derivative of 4-FMC. Fig. 9D shows the TIC of the mixture of the PFPA derivatives (Fluoro PFPA Mix). An expanded chromatogram can be seen in Fig. A22. 4-FMC-PFPA elutes at 1.997 minutes, 3-FMC-PFPA elutes at 2.054 minutes, and 2-FMC-PFPA elutes at 2.376 minutes (mass spectra in Figs. A23-A25, respectively). The mass spectra for the PFPA derivatives are found in Fig. 10. The three mass spectra contain the same major ions, but, like the TFAA and HFBA derivatives, the ions are in different relative ratios. This difference in ratios allows the isomers to be easily differentiated and identified. The major ions and their ion ratios compared to the base peak can be found in Table 3



 $\frac{1}{250}$ $\frac{1}{450}$ $\frac{1}{450}$ $\frac{1}{450}$ $\frac{1}{100}$ $\frac{1}$



Fig. 10: Mass spectra of PFPA Derivatives of the Fluoromethcathinones. A: 2-FMC-PFPA; B: 3-FMC-PFPA; C: 4-FMC-PFPA.

While the Fluoromethcathinones can be easily distinguished on their own by retention time, the same is not true of their mass spectra. 2-FMC is differentiable from 3-FMC and 4-

FMC. However, the mass spectra of 3- and 4-FMC are nearly identical with only a small amount of difference in the low abundance ions. Derivatization using TFAA was successful in differentiating the mass spectral results, but 3-FMC and 4-FMC co-eluted with TFAA derivatization. Both TFAA and HFBA were able to eliminate all breakdown compounds. HFBA was successful in achieving both chromatographic separation of the isomers and in differentiating the mass spectra so that all three isomers can be identified. PFPA also succeeded in chromatographically separating the isomer and differentiating the mass spectra making identification possible. On the other hand, PFPA was not able to eliminate all breakdown products. Because HFBA was successful in all three categories, it is considered the best choice for derivatizing the Fluoromethcathinones.

Methoxymethcathinones

The TICs for 2-Methoxymethcathinone (2-MOMC), 3-Methoxymethcathinone (3-MOMC), Methedrone (4-MOMC), and the Methoxymethcathinone mixture (Methoxy Standard Mix) are shown in Fig. 11. The TIC for 2-MOMC (Fig. 11A) includes four peaks. 2-MOMC decomposes in the GC-MS like 2-FMC. The peak at 2.904 minutes is breakdown (mass spectrum in Appendix B Fig. B1). 2-MOMC elutes at 4.454 minutes. The peaks at 4.622 and 4.643 minutes have nearly identical mass spectra, both of which correspond with the 2-MOMC enamine (Figs. B2 and B3, respectively). The 3-MOMC chromatogram (Fig. 11B) shows only one peak, at 5.125 minutes, that of 3-MOMC. Similarly, the Methedrone TIC (Fig. 11C) also only shows Methedrone eluting at 6.866 minutes. Fig. 11D presents the TIC for the Methoxy standard mixture, and an expanded view of this chromatogram can be seen in Fig. B4. Figs. B5-B7 show the mass spectra for this mixture; 2-MOMC elutes at 4.415 minutes, 3-MOMC elutes at 5.104 minutes, and Methedrone elutes at 6.841 minutes. The peaks at 2.358, 2.580, and 2.897 minutes are all breakdown and rearrangement compounds observed when the standards were run. The mass spectra for the Methoxymethcathinone standards are in Fig. 12. Table 4 gives the major ions found in each mass spectrum along with the ion ratios.

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The mass spectra for the positional isomers of the MOMC compounds are more easily differentiated than the three Fluoromethcathinone positional isomers, but are still substantially similar enough that it would be difficult to differentiate between them by GC-MS alone.



Fig. 11: Total Ion Chromatograms of the Methoxymethcathinone Standards. A: 2-MOMC; B: 3-MOMC; C: Methedrone; D: Methoxy Standard Mix.



Fig. 12: Mass spectra of the Methoxymethcathinone Standards. A: 2-MOMC; B: 3-MOMC; C: Methedrone.

Compound	Retention	Base Peak	Major Ions in m/z (Base Peak
_	Time	(Abundance)	Abundance/Ion Abundance Ratio)
2~MOMC	4.54 min	58 m/z	42(38), 77(10), 92(21), 105(125),
		(312725)	121(92), 135(14), 193(164)
3~MOMC	5.12 min	58 m/z	42(36), 77(15), 92(16), 107(30),
		(133386)	135(23), 193(745)
Methedrone	6.86 min	58 m/z	42(40), 77(15), 92(16), 107(48),
		(325985)	135(11), 1933190
2~MOMC~TFAA	6.66 min	135 m/z	42(32), 56(57), 77(9.7), 92(91),
		(610816)	110(20), 136(11), 154(13), 289(270)
3-MOMC-TFAA	6.76 min	135 m/z	42(16), 56(36), 77(10), 92(12),
		(474112)	107(6.7), 110(7.9), 136(11), 154(2.4),
			289(21)
Methedrone-TFAA	7.99 min	135 m/z	42(15), 56(72), 77(13), 92(16), 107(21),
		(1077248)	110(22), 136(11), 154(14), 289(95)
2~MOMC~HFBA	5.94 min	135 m/z	42(31), 56(44), 66(1578), 77(10),
		(755968)	92(22), 105(104), 120(90), 136(11),
			169(34), 210(15), 254(8.3), 389(1035)
3~MOMC~HFBA	6.39 min	135 m/z	42(15), 56(24), 66(658), 77(10), 92(13),
		(719616)	107(7.6), 119(97), 136(11), 169(16),
			210(4.6), 254(1.4), 389(25)
Methedrone-HFBA	7.59min	135 m/z	42(33), 56(55), 66(808), 77(14), 92(18),
		(1620992)	107(24), 119(179), 136(11), 169(36),
			210(16), 254(8.5), 389(230)
2-MOMC-PFPA	5.79 min	135 m/z	42(29), 56(47), 64(77), 77(9.9), 92(20),
		(1085952)	105(103), 119(24), 136(11), 160(15),
			176(59), 204(10), 339(692)
3-MOMC-PFPA	6.10 min	135 m/z	42(14), 56(28), 64(27), 77(10), 92(12),
		(631872)	107(7.3), 119(13), 136(11), 160(5.1),
			176(35), 204(1.7), 339(24)
Methedrone~PFPA	7.54 min	135 m/z	42(31), 56(58), 64(44), 77(13), 92(16),
		(2530304)	107(21), 119(25), 136(10), 160(16),
			176(53), 204(9.7), 339(156)

Table 4: Methoxymethcathinone Data

The TICs for the TFAA derivatives of the Methoxymethcathinones are displayed in Fig. 13. The retention times for the TFAA derivatives of the Methoxymethcathinones can be found in Table 4. Fig. 13D gives the TIC for the Methoxy TFAA mixture (Methoxy TFAA Mix). An expanded view of this chromatogram is shown in Fig. B8. 2-MOMC-TFAA elutes at 6.652 minutes, 3-MOMC-TFAA elutes at 6.754 minutes, and Methedrone-TFAA elutes at 7.991 minutes (mass spectra in Figs. B9-B11, respectively). Fig. 14 presents the mass spectra of the TFAA derivatives of the Methoxymethcathinones. The major ions of the derivatives and their ion ratios can be seen in Table 4.



Fig. 13: Total Ion Chromatograms of TFAA Derivatives of the Methoxymethcathinones. A: 2^{-7} MOMC-TFAA; B: 3-MOMC-TFAA; C: Methedrone-TFAA; D: Methoxy TFAA Mix.



B: 3-MOMC-TFAA; C: Methedrone-TFAA.

Fig. 15 presents the TICs for the HFBA derivatives of the Methoxymethcathinones. The retention times of the HFBA derivatives are found in Table 4. The mixture of these derivatives is displayed in Fig. 15D (Methoxy HFBA Mix). 2-MOMC-HFBA elutes at 5.897 minutes, 3MOMC-HFBA elutes at 6.350 minutes, and Methedrone-HFBA elutes at 7.579 minutes (mass spectra in Figs. B12-B14, respectively). The mass spectra for the HFBA derivatives are given in Fig. 16. The major ions of the Methoxymethcathinone HFBA derivatives and their ion ratios are given in Table 4.



Fig. 15: Total Ion Chromatograms of HFBA Derivatives of the Methoxymethcathinones. A: 2~ MOMC~HFBA; B: 3~MOMC~HFBA; C: Methedrone~HFBA; D: Methoxy HFBA Mix.



B: 3-MOMC-HFBA; C: Methedrone-HFBA.

The TICs for the PFPA derivatives of the Methoxymethcathinones are shown in Fig. 17. Fig. 17A contains 2 peaks; the peak at 5.795 minutes is 2-MOMC-PFPA, and the peak at 10.624 minutes is the enamine of the derivative (mass spectrum shown in Fig. B15). Fig. 17B shows one peak at 6.104 minutes which corresponds to 3-MOMC-PFPA. Fig. 17C contains two peaks. The first peak at 7.542 minutes is Methedrone-PFPA. The second peak at 9.892 minutes is breakdown of the compound in the GC (mass spectrum can in Fig. B16). The mixture of these derivatives is seen in Fig. 17D (Methoxy PFPA Mix). 2-MOMC-PFPA elutes at 5.727 minutes, 3-MOMC-PFPA elutes at 6.041 minutes, and Methedrone-PFPA elutes at 7.501 minutes (mass spectra in Figs. B17-B19, respectively). Fig. 18 displays the mass spectra of the PFPA derivatives of the Methoxymethcathinones. Table 4 gives the major ions of these PFPA derivatives and their ion ratios.



Fig. 17: Total Ion Chromatograms of PFPA Derivatives of the Methoxymethcathinones. A: 2-MOMC-PFPA; B: 3-MOMC-PFPA; C: Methedrone-PFPA; D: Methoxy PFPA Mix.



Fig. 18: Mass spectra of PFPA Derivatives of the Methoxymethcathinones. A: 2-MOMC-PFPA; B: 3-MOMC-PFPA; C: Methedrone-PFPA.

The Methoxymethcathinones are identifiable by their retention times. The mass spectrum of 2-MOMC is easily differentiated from 3-MOMC and Methedrone. The mass spectra of 3-MOMC and Methedrone contain the same ions, with only one difference, in almost the same ratios. TFAA was once again successful in differentiating the mass spectra of the isomers and in preventing breakdown compounds. However, TFAA was not able to completely resolve 2-MOMC and 3-MOMC in the mixture. HFBA was successful in all three categories: the compounds are easily separated when in a mixture, all breakdown products were prevented, and the mass spectra are easily differentiated. PFPA was able to separate the compounds chromatographically and in creating mass spectra that could be differentiated. However, PFPA was not successful in preventing all breakdown products. Given that HFBA was successful in all three areas, it would be the best choice for derivatization of the Methoxymethcathinones.

Methylmethcathinones

The TICs of 2-Methylmethcathinone (2-MMC), 3-Methylmethcathinone (3-MMC), Mephedrone (4-MMC), and the Methylmethcathinone mixture (Methyl Standard Mix) are displayed in Fig. 19. The TIC for 2-MMC (Fig. 19A) contains four peaks. The peaks at 1.228 and 1.380 minutes are either breakdown or rearrangements (their respective mass spectra are found in Appendix C Figs. C1 and C2). The peaks at 2.073 and 2.353 minutes are 2-MMC; the high sample concentration leads to some 2-MMC eluting later. Fig. 19B shows the chromatogram for 3-MMC with one peak at 2.532 identified as 3-MMC. Mephedrone's chromatogram is seen in Fig. 19C and has two peaks. The first peak at 2.843 minutes is that of Mephedrone and the peak at 3.155 minutes is the enamine of Mephedrone (mass spectrum in Fig. C3). Fig 19D gives the TIC for the Methyl standard mixture. An expanded chromatogram of this mix can be seen in Fig. C4. Due to the low concentration of the sample, column bleed is observed at 0.901, 0.999, 1.706, and 11.870 minutes. The peaks at 1.224 and 1.312 minutes are breakdown that were seen when the standards were run. The peak at 2.070 minutes is 2-MMC, the peak at 2.518 minutes is 3-MMC, and the peak at 2.812 minutes is Mephedrone (mass spectra in Figs. C5-C7, respectively). The similarity of the Methylmethcathinone mass spectra can be observed in Fig. 20. All three mass spectra contain the same major ions in the

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same relative abundance. Table 5 gives the major ions of the Methylmethcathinones and their ion ratios compared to their base peaks.



Fig. 19: Total Ion Chromatograms of the Methylmethcathinone Standards. A: 2-MMC; B: 3-MMC; C: Mephedrone; D: Methyl Standard Mix



Fig. 20: Mass spectra of the Methylmethcathinone Standards. A: 2-MMC; B: 3-MMC; C: Mephedrone.

Table 5: Methylmethcathinor	ie Data
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Compound	Retention	Base Peak	Major Ions in m/z (Base Peak
_	Time	(Abundance)	Abundance/Ion Abundance Ratio)
2~MMC	2.07 min	58 m/z	42(33), 65(14), 77(105), 91(9.4),
		(236262)	105(127), 119(21), 177(865)
2-MMC	2.53 min	58 m/z	42(35), 65(15), 77(138), 91(10),
		(248056)	105(160), 119(24), 177(515)
Mephedrone	2.84 min	58 m/z	42(37), 65(15), 77(139), 91(10),
		(1434624)	105(142), 119(19), 177(346)
2-MMC-TFAA	3.58 min	119 m/z	42(21), 56(40), 65(14), 91(3.8),
		(233728)	110(11), 120(11), 154(5.1), 273(237)
3-MMC-TFAA	3.83 min	119 m/z	42(18), 56(38), 65(14), 91(4.1),
		(1119232)	110(8.6), 120(11), 154(2.9), 273(62)
Mephedrone-TFAA	4.27 min	119 m/z	42(22), 56(50), 65(16), 91(4.7),
		(625280)	110(12), 120(11), 154(5.1), 273(97)
2-MMC-HFBA	3.24 min	119 m/z	42(21), 56(35), 65(16), 77(181),
		(1110016)	91(83), 120(11), 169(23), 210(8.4),
			254(3.0), 373(421)
3-MMC-HFBA	3.32 min	119 m/z	42(17), 56(28), 65(15), 77(197),
		(1284608)	91(4.4), 120(11), 169(17), 210(5.4),
			254(1.6), 373(118)
Mephedrone-HFBA	3.77 min	119 m/z	42(20), 56(35), 65(18), 77(254),
		(1111040)	91(5.5), 120(11), 169(22), 210(8.1),
	-		254(2.8), 373(229)
2-MMC-PFPA	3.12 min	119 m/z	42(20), 56(40), 65(16), 77(185),
		(1461832)	91(4.3), 120(11), 160(7.6), 204(3.9),
			323(333)
3-MMC-PFPA	3.15 min	119 m/z	42(16), 56(33), 65(16), 77(199),
		(1423360)	91(4.5), 120(12), 160(5.7), 204(2.2),
			323(91)
Mephedrone~PFPA	3.63 min	119 m/z	42(21), 56(40), 65(17), 77(254),
		(1849856)	91(5.4), 120(11), 160(7.6), 204(3.6),
			323(171)

The TICs of the TFAA derivatives of the Methylmethcathinones are provided in Fig. 21. The retention times for the TFAA derivatives are seen in Table 5. The TIC for the Methylmethcathinone TFAA mixture (Methyl TFAA Mix) is provided in Fig. 21D. This mixture contains only two peaks. The first peak at 3.169 minutes is the co-elution of 2-MMC-TFAA and 3-MMC-TFAA (mass spectrum in Fig. C8) and the second peak at 4.261 minutes is Mephedrone-TFAA (mass spectrum in Fig. C9). The mass spectra of the TFAA derivatives of the Methylmethcathinones are presented in Fig. 22. Table 4 provides the major ions and their ion ratios.



Fig. 21: Total Ion Chromatograms of TFAA Derivatives of the Methylmethcathinones. A: 2-MMC-TFAA; B: 3-MMC-TFAA; C: Mephedrone-TFAA; D: Methyl TFAA Mix.



Fig. 22: Mass spectra of TFAA Derivatives of the Methylmethcathinones. A: 2-MMC-TFAA; B: 3-MMC-TFAA; C: Mephedrone-TFAA.

The TICs of the HFBA derivatives of the Methylmethcathinones are shown in Fig. 23. 2-MMC-HFBA elutes at 3.247 minutes (Fig. 23A) and is the only compound observed in that chromatogram. Conversely, Fig. 23B shows two peaks at 2.525 and 3.325 which are a small amount of underivatized 3-MMC (mass spectrum in Fig. C10), and 3-MMC-HFBA, respectively. Mephedrone-HFBA elutes at 3.776 minutes and is the only peak in its chromatogram (Fig. 23C). Fig. 23D shows the TIC for the HFBA mixture of the Methyls (Methyl HFBA Mix). Fig. C11 shows an expanded chromatogram of the mixture. 2-MMC-HFBA elutes at 3.239 minutes, 3-MMC-HFBA elutes at 3.313 minutes, and Mephedrone-HFBA elutes at 3.752 minutes (mass spectra in Figs. C12-C14, respectively). The mass spectra of the Methyl HFBA derivatives are seen in Fig. 24. The major ions and their ratios are found in Table 5.



Fig. 23: Total Ion Chromatograms of HFBA Derivatives of the Methylmethcathinones. A: 2-MMC-HFBA; B: 3-MMC-HFBA; C: Mephedrone-HFBA; D: Methyl HFBA Mix.



Fig. 24: Mass spectra of HFBA Derivatives of the Methylmethcathinones. A: 2-MMC-HFBA; B: 3-MMC-HFBA; C: Mephedrone-HFBA.

Fig. 25 gives the TICs for the PFPA derivatives of the Methylmethcathinones. Fig. 25A shows the TIC for 2-MMC-PFPA; 2-MMC-PFPA elutes at 3.123 minutes and the peak at 9.801 minutes is column breakdown (mass spectrum in Fig. C15). 3-MMC-PFPA elutes at 3.155 minutes and is the only compound in the corresponding TIC (Fig. 25B). The chromatogram in Fig. 25C contains three peaks. The first two peaks at 2.826 and 3.128 minutes are breakdown and their mass spectra can be seen in Figs. C16 and C17. The peak at 3.635 minutes is Mephedrone-PFPA. The TIC for the PFPA Methyl mixture (Methyl PFPA Mix) is seen in Fig. 25D. This chromatogram has two peaks even though there should be three separate compounds. The first peak, at 3.146 minutes, is the co-elution of 2-MMC-PFPA and 3-MMC-PFPA. The second peak at 3.618 minutes is Mephedrone-PFPA. Mass spectra of the PFPA derivatives are displayed in Fig. 26. The major ions and their rations can be found in Table 5.



Fig. 25: Total Ion Chromatograms of PFPA Derivatives of the Methylmethcathinones. A: 2-MMC-PFPA; B: 3-MMC-PFPA; C: Mephedrone-PFPA; D: Methyl PFPA Mix.



Fig. 26: Mass spectra of PFPA Derivatives of the Methylmethcathinones. A: 2-MMC-PFPA; B: 3-MMC-PFPA; C: Mephedrone-PFPA.

The Methylmethcathinones are differentiated by their retention times. The major ions seen are the same for the isomers with some difference seen amongst the low abundance ions. The use of TFAA was successful in eliminating breakdown product, but chromatographic

separation of the compounds was not achieved. Further, the mass spectra showed only slight differences in the relative abundance of the various ions. The use of HFBA was successful in partially eliminating breakdown products. HFBA derivatives of the isomers separated chromatographically and showed better mass spectral results. *3-MMC* is easily distinguishable when not derivatized, but 2-MMC and Mephedrone are better distinguished when derivatized with HFBA. Derivatization with PFPA, like TFAA, did not result in chromatographic separation of the isomers and, like HFBA, did not eliminate breakdown products. PFPA derivatization allowed for the easy differentiation of *3-MMC* from 2-MMC and Mephedrone. While underivatized 2-MMC and Mephedrone can be distinguished from one another, it is much more difficult than with *3-MMC*. None of the derivatizing agents were successful in all three areas, but as HFBA was the only agent to lead to separation of the isomers and would be the best choice for derivatization of the Methylmethcathinones.

Methylenedioxymethcathinones

The TICs of 2,3-Methylenedioxymethcathinone (2,3-MDMC), Methylone (3,4-MDMC), and the Methylenedioxy standard mixture (MD Standard Mix) are shown in Fig. 27. The TIC in Fig. 27A contains three peaks. The first peak at 7.549 minutes is 2,3-MDMC; the peak at 7.878 minutes is the enamine of 2,3-MDMC and the mass spectrum is located in Appendix D Fig. D1; the final peak at 17.513 minutes is either air or cap bleed (mass spectrum not shown). Fig. 27B is the TIC for Methylone showing two peaks. The peak at 8.175 minutes is Methylone and the peak at 8.472 minutes is the enamine of Methylone, whose mass spectrum is found in Fig. D2. The TIC of the MD Standard mix is shown in Fig. 27C. Many peaks are observed in this chromatogram. Due to the low sample concentration in the mixture, the amount of column and/or vial cap bleed seen is much higher and more noticeable. Peaks at 0.999, 1.788, 30465, 6.692, 8.348 minutes, and later are column bleed or cap bleed and are not relevant. An expanded TIC can be viewed in Fig. D3. The peaks at 5.031 and 7.878 are breakdown components of 2,3-MDMC and Methylone. 2,3-MDMC

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elutes at 7.524 minutes (mass spectrum in Fig. D4) and Methylone elutes at 8.164 minutes (mass spectrum in Fig. D5). The similarity between the mass spectra of 2,3-MDMC and Methylone is seen in Fig. 28. The major ions of the Methylenedioxymethcathinones and their ion ratios are found in Table 6.



Fig. 27: Total Ion Chromatograms of the Methylenedioxymethcathinone Standards. A: 2,3~ MDMC; B: Methylone; C: MD Standard Mix



Fig. 28: Mass spectra of the Methylenedioxymethcathinone Standards. A: 2,3-MDMC; B: Methylone.

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Compound	Retention	Base Peak	Major Ions in m/z (Base Peak
	Time	(Abundance)	Abundance/Ion Abundance Ratio)
2,3~MDMC	7.54 min	58 m/z	42(33), 65(13), 91(34), 121(30),
		(778368)	135(108), 149(16), 207(134)
Methylone	8.17 min	58 m/z	42(43), 64(17), 91(35), 121(17),
		(623232)	135(107), 149(13), 207(262)
2,3-MDMC-TFAA	8.71 min	149 m/z	42(17), 56(35), 65(9.8), 91(25),
		(2281472)	110(7.6), 121(37), 150(10),
			152(2962), 160(65), 178(60), 303(8.1)
Methylone-TFAA	8.97 min	149 m/z	42(27), 56(55), 65(17), 91(35),
		(2083840)	110(14), 121(9.3), 150(11), 152(729),
			160(164), 178(529), 303(16)
2,3~MDMC~HFBA	8.34 min	149 m/z	42(16), 56(24), 65(10), 91(24),
		(2391552)	121(38), 150(10), 169(15), 210(4.1),
			254(1.3), 403(7.9)
Methylone~HFBA	8.68 min	149 m/z	42(28), 56(42), 65(177), 91(34),
		(3383808)	121(10), 150(11), 169(26), 210(9.5),
			254(4.1), 403(20)
2,3~MDMC~PFPA	8.28 min	149 m/z	42(15), 56(27), 65(10), 91(26),
		(1756160)	119(12), 150(11), 160(4.5), 178(50),
			204(1.8), 353(7.6)
Methylone~PFPA	8.63 min	149 m/z	42(25), 56(45), 65(17), 91(34),
		(3144192)	121(9.8), 150(11), 160(9.4), 178(401),
			204(5.2), 353(16)

Table 6: Methylenedioxymethcathinone Data

Fig. 29 shows the TICs of the Methylenedioxy standards derivatized with TFAA. 2,3-MDMC-TFAA is seen in Fig. 29A at 8.716 minutes. This chromatogram contains another peak at 10.289 minutes; this is a breakdown or rearrangement product, the mass spectrum of which can be seen in Fig. D6. Methylone-TFAA elutes at 8.977 minutes and can be seen in Fig. 29B. The Methylenedioxy mixture of these derivatives (MD TFAA Mix) is seen in Fig. 29C. There are three peaks within the chromatogram. The peaks at 8.712 and 8.978 minutes are 2,3-MDMC-TFAA and Methylone-TFAA, respectively. The mass spectra of these can be seen in Figs. D7 and D8. The peak at 10.286 minutes is the breakdown/rearrangement product of 2,3-MDMC-TFAA. The mass spectra of the TFAA derivatives can be seen in Fig. 30. The major ions and their ion ratios can be seen in Table 6.



Fig. 29: Total Ion Chromatograms of TFAA Derivatives of the Methylenedioxymethcathinones. A: 2,3-MDMC-TFAA; B: Methylone-TFAA; C: MD TFAA Mix.



Fig. 30: Mass spectra of TFAA Derivatives of the Methylenedioxymethcathinones. A: 2.3-MDMC+TFAA; B: Methylone-TFAA.

The TICs of the HFBA derivatives of the Methylenedioxy compounds are shown in Fig. 31. Fig. 31A shows a chromatogram with two peaks. The first is 2,3-MDMC-HFBA at 8.346 minutes and the second, at 10.056 minutes, is a breakdown product (mass spectrum in Fig. D9). Fig. 31B shows the chromatogram of Methylone-HFBA which elutes at 8.686 minutes and a breakdown product at 11.559 minutes (mass spectrum in Fig. D10). The TIC for the mixture of the HFBA derivatives (MD HFBA Mix) is found in Fig. 31C. This chromatogram contains four peaks. At 8.341 minutes is 2,3-MDMC-HFBA and at 8.674 minutes is Methylone-HFBA (mass spectra in Fig. D11 and D12, respectively). At 10.057 minutes is the

breakdown from 2,3-MDMC-HFBA and at 11.332 minutes is the breakdown from Methylone-HFBA. Fig. 32 displays the mass spectra of the HFBA derivatives of the Methylenedioxy positional isomers. The major ions of the HFBA derivatives and their ratios can be found in Table 6.



Fig. 31: Total Ion Chromatograms of HFBA Derivatives of the Methylenedioxymethcathinones. A: 2,3-MDMC-HFBA; B: Methylone-HFBA; C: MD HFBA Mix.



MDMC+HFBA; B: Methylone~HFBA

Total ion chromatograms of the PFPA derivatives of the methylenedioxy isomers are shown in Fig. 33. The TIC of 2,3-MDMC-PFPA is found in Fig. 33A. This TIC contains two peaks: the one at 8.282 minutes is 2,3-MDMC-PFPA and at 10.075 minutes is a breakdown product (mass spectrum in Fig. D13). Methylone-PFPA elutes at 8.633 minutes and is seen in Fig. 33B. The TIC of the mixture of the PFPA derivatives (MD PFPA Mix) is in Fig. 33C. An expanded view of this chromatogram is in Fig. D14. The peak at 8.277 minutes is 2,3-MDMC-PFPA (mass spectrum in Fig. D15), the peak at 8.619 minutes is Methylone-PFPA (mass spectrum in Fig. D16), and the peaks at 10.076 and 11.333 minutes are both breakdown products of 2,3-MDMC and Methylone. The mass spectra of the PFPA derivatives are shown in Fig. 34. The major ions of each spectrum and their ion ratios compared to the base peak of the PFPA derivatives are given in Table 6.



Fig. 33: Total Ion Chromatograms of PFPA Derivatives of the Methylenedioxymethcathinones. A: 2,3-MDMC-PFPA; B: Methylone-PFPA; C: MD PFPA Mix.



Fig. 34: Mass spectra of PFPA Derivatives of the Methylenedioxymethcathinones. A: 2.3-MDMC+PFPA; B: Methylone-PFPA.

Differentiation of the Methylenedioxymethcathinones is possible based on retention times. However, with the exception of the abundance of one ion, the mass spectra are nearly identical. Derivatization with TFAA was allowed for the separation of the compounds within a mixture, but not prevent breakdown. TFAA derivatization made differentiation of the mass spectral results possible. HFBA derivatization led to separation of the isomers and provided differentiation between the mass spectra, but did not prevent breakdown from occurring. PFPA derivatization allowed for chromatographic separation but did not prevent breakdown of the Methylenedioxymethcathinones. An advantage to using PFPA for derivatization was that it led to different relative abundance ratios and an identifying ion for each of the isomers. All three derivatizing agents led to the successful separation of the isomers and their successful differentiation, but did not prevent breakdown. Because PFPA was the only derivatizing agent to lead to a difference between the major ions, it would likely be the best option for the derivatization of the Methylenedioxymethcathinones; although, HFBA and TFAA would also lead to the successful differentiation of the MDMCs. Table 7 demonstrates the advantages and disadvantages of each perfluoroacyl anhydride when used to derivatize the synthetic cathinones with positional isomers.

Group	TFAA	HFBA	PFPA
Fluoro~	Co-elution in a	Compounds	Compounds
methcathinones	mixture; prevented	separated in a	separated in a
	breakdown;	mixture; prevented	mixture; some
	differentiated MS	breakdown;	breakdown seen;
		differentiated MS	differentiated MS
Methoxy~	Compounds not fully	Compounds	Compounds
methcathinones	resolved in a mixture;	separated in a	separated in a
	prevented	mixture; prevented	mixture; some
	breakdown;	breakdown;	breakdown seen;
	differentiated MS	differentiated MS	differentiated MS
Methyl~	Co-elution in a	Compounds	Co-elution in a
Methcathinones	mixture; prevented	separated in a	mixture; some
	breakdown; slight	mixture; some	breakdown seen; 3~
	differences in MS	breakdown seen; 3-	MMC MS easily
		MMC MS easily	distinguishable, slight
		distinguishable, slight	difference between 2-

Table 7: Advantages and Disadvantages of TFAA, HFBA, and PFPA

		difference between 2- MMC and	MMC and Mephedrone MS
		Mephedrone MS	
Methylenedioxy~	Compounds	Compounds	Compounds
methcathinones	separated in a	separated in a	separated in a
	mixture; some	mixture; some	mixture; some
	breakdown seen;	breakdown seen;	breakdown seen;
	differentiated MS	differentiated MS	differentiated MS
			with 2,3~MDMC
			having an ion at 119
			m/z and Methylone
			having an ion @ 121
			m/z

Cathinones Containing a Pyrrolidine

The total ion chromatograms of the Pyrrolidinopropiophenone standards are presented in Fig. 35. Alpha-Pyrrolidinopropiophenone (PPP) elutes at 6.317 minutes (Fig. 35A). The peak observed at 7.282 minutes is either air or cap bleed (mass spectrum not shown). 2-Methyl-αpyrrolidinopropiophenone (2-MPPP) elutes at 7.314 minutes (Fig. 35B). There are also three other peaks observed in this TIC: the peaks at 0.899 and 1.233 minutes are both breakdown (mass spectrum in Appendix E Fig. E1 and E2). The peak observed at 8.595 minutes is the enamine of 2-MPPP (mass spectrum in Fig. E3). 3-Methyl-α-pyrrolidinopropiophenone (3-MPPP) elutes at 7.880 minutes (Fig. 35C) and its enamine at 9.036 minutes (mass spectrum in Fig. E4). 4-Methyl-α-pyrrolidinopropiophenone (4-MPPP) elutes at 8.233 minutes (Fig. 35D). There are two additional peaks in this mass spectrum; the peak at 7.845 minutes is breakdown (mass spectrum in Fig. E5) and the peak at 9.285 minutes is the enamine of 4-MPPP (mass spectrum in Fig. E6). 4-Methoxy- α -pyrrolidinopropiophenone (4-MOPPP) elutes at 9.795 minutes (Fig. 35E) and its enamine at 10.332 minutes (mass spectrum in Fig. E7). 3,4-Methylenedioxy-α-pyrrolidinopropiophenone (3,4-MDPPP) elutes at 10.265 minutes (Fig. 35F) and its enamine at 10.710 minutes (mass spectrum in Fig. E8). Derivatization of these compounds was unsuccessful due to the presence of a tertiary nitrogen and subsequently no

mixture analysis was performed on these compounds. The mass spectra of the

Pyrrolidinopropiophenones are shown in Fig. 36. They each contain the base peak of 98 m/z. The mass spectra of 2-MPPP, 3-MPPP, and 4-MPPP are nearly identical, as they are positional isomers, making differentiation difficult. However, it is possible to distinguish the mass spectra of the Methyl-PPPs from PPP, 4-MOPPP, and 3,4-MDPPP.



Fig. 35: Total Ion Chromatograms of the Pyrrolidinopropiophenone Standards. A: α-PPP; B: 2-MPPP; C: 3-MPPP; D: 4-MPPP; E: 4-MOPPP; F: 3,4-MDPPP.

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Fig. 36: Mass spectra of the Pyrrolidinopropiophenone Standards. A: α-PPP; B: 2-MPPP; C: 3-MPPP; D: 4-MPPP; E: 4-MOPPP; F: 3,4-MDPPP.

The TICs of Pyrovalerone and Methylenedioxypyrovalerone (MDPV) are provided in Fig. 37. Fig. 37A shows Pyrovalerone has a retention time of 9.283 minutes with the enamine eluting at 9.695 minutes (mass spectrum in Fig. E9). Fig. 37B shows MDPV eluting at 10.678 minutes and the enamine at 10.905 minutes (mass spectrum in Fig. E10). Derivatization on these two compounds was also unsuccessful because they contain a tertiary nitrogen. The mass spectra of Pyrovalerone and MDPV (Fig. 38) are similar. They both contain a base peak at 126 m/z, but the other major ions are different allowing for differentiation of the compounds.



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Other Cathinones

Cathinone, Pentedrone, Butylone, and 3,4-Dimethylmethcathinone do not have any positional isomers that were tested in this study; however, the data for these four compounds is presented in the event that an isomer is developed or becomes abused on the illicit market. The TICs for Cathinone and its perfluoroacyl anhydrides are shown in Fig. 39. Cathinone has a retention time of 1.655 minutes (Fig. 39A), Cathinone-TFAA of 2.159 minutes (Fig. 39B), Cathinone-HFBA of 2.019 minutes (Fig. 39C), and Cathinone-PFPA of 1.897 minutes (Fig. 39D). The mass spectra for the Cathinones can be seen in Fig. 40. Cathinone has a base peak of 44 m/z, while all the derivatized Cathinone compounds have a base peak of 105 m/z. The mass spectra are easily distinguished when the other ions are considered.



Fig. 39: Total Ion Chromatograms of the Cathinone Standards. A: Cathinone; B: Cathinone-TFAA; C: Cathinone-HFBA; D: Cathinone-PFPA.



Fig. 40: Mass spectra of the Cathinone Standards. A: Cathinone; B: Cathinone-TFAA; Cathinone-HFBA; D: PFPA.

The chromatograms for Pentedrone and its derivatives are found in Fig. 41. Pentedrone elutes at 3.048 minutes (Fig. 41A), Pentedrone-TFAA at 4.306 minutes (Fig. 41B), Pentedrone-HFBA at 3.783 minutes (Fig. 41C), and Pentedrone-PFPA at 3.596 minutes (Fig. 41D). The

mass spectra for the Pentedrones are in shown in Fig. 42. Pentedrone has a base peak at 86 m/z; Pentedrone-TFAA at 182 m/z; Pentedrone-HFBA at 282 m/z; and Pentedrone-PFPA at 232 m/z. These are all easily distinguished from one another having different base peaks and different ions.



Fig. 41: Total Ion Chromatograms of the Pentedrone Standards. A: Pentedrone; B: Pentedrone-TFAA; C: Pentedrone-HFBA; D: Pentedrone-PFPA.



Pentedrone-HFBA; D: Pentedrone-PFPA.

The TICs for Butylone and its derivatives are given in Fig. 43. Butylone elutes at 8.798 minutes (Fig. 43A), Butylone-TFAA at 9.287 minutes (Fig. 43B), Butylone-HFBA at 8.985 minutes (Fig. 43C), and Butylone-PFPA at 8.965 minutes (Fig. 43D). The mass spectra for the Butylones are shown in Fig. 44. Butylone has a base peak of 72 m/z, while all the derivatized Butylone compounds have a base peak at 149 m/z. However, looking at the other ions, the mass spectra are still easily distinguished from one another.



Fig. 43: Total Ion Chromatograms of the Butylone Standards. A: Butylone; B: Butylone-TFAA; C: Butylone-HFBA; D: Butylone-PFPA.



Fig. 44: Mass spectra of the Butylone Standards: A:Butylone; B: Butylone-TFAA; C: Butylone-HFBA; D: Butylone-PFPA.

The chromatograms for 3,4-Dimethylmethcathinone (3,4-diMMC) and its derivatives are displayed in Fig. 45. 3,4-diMMC at 5.099 minutes (Fig. 45A), 3,4-diMMC-TFAA at 6.863 minutes (Fig. 45B), 3,4-diMMC-HFBA at 6.309 minutes (Fig. 45C), and 3,4-diMMC-PFPA at

6.039 minutes (Fig. 45D). The mass spectra of 3,4-diMMC and its derivatives can be seen in Fig. 46. 3,4-diMMC has a base peak of 58 m/z, while the derivatives of 3,4-diMMC contain a base peak of 133 m/z. The other ions in the mass spectra are sufficiently different that differentiation is possible.



diMMC-TFAA; C: 3,4-diMMC-HFBA; D: 3,4-diMMC-PFPA.



3,4-diMMC-HFBA; D: 3,4-diMMC-PFPA.

Conclusions

The mass spectra of the TFAA, HFBA, and PFPA derivatives of 15 synthetic cathinones were studied and compared to one another and those of the underivatized compounds. The presence of a tertiary nitrogen prevented the derivatization of eight standards because perfluoroacyl anhydrides can only derivatize primary and secondary nitrogens. Pyridine was used in an attempt to enolize the carbonyl and open the pyrrolidine ring for derivatization, but this method was unsuccessful. HFBA derivatization allowed for the differentiation between the mass spectra of positional isomers better than TFAA or PFPA, overall. PFPA was more successful than HFBA or TFAA for the derivatization of the Methylenedioxymethcathinones isomers. When choosing between the three derivatizing agents studied, HFBA would give the best overall results as it was successful in differentiating all of the positional isomers. The Future studies will include the reproduction of the current results, analysis of more complex cathinone mixtures, and development of a derivatization method for the compounds containing tertiary nitrogens.

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References

- 1. Federal Analogue Act 21 U.S.C. § 813.
- S3187 amendment of the Federal Food, Drug, and Cosmetic Act (21U.S.C. 301) Enacted July 9, 2012 http://www.gpo.gov/fdsys/pkg/BILLS-112s3187enr/pdf/BILLS-112s3187enr.pdf.
- 3. Archer RP. *Fluoromethcathinone, a new substance*. Forensic Science International. 2009; 185: 10-20.
- LaGrone E, Kiyak C, Rodriguez G, Rankin JG. *Trifluoroacetyl Derivatization of Amphetamine, Methamphetamine, MDMA and Other Controlled Substances* with Similar Mass spectra. American Academy of Forensic Science: Proceedings. 2011; 17(A3): 23.
- Clark CR, Abdel-Hay KM, Awad T, DeRuiter J. Differentiation of methylenedioxybenzylpiperazines (MDBFs) and methoxymethylbenzylpiperazines (MMBFs) by GC-IRD and GC-MS. Forensic Science International. 2011; 210: 122-128.
- Lum BJ, Hibbert DB, Brophy J. Identification of Substituted Cathinones (β-keto phenethylamines) by Heptafluorobutyric Anhydride (HFBA) Chemical Derivatization and Gas Chromatography Mass Spectrometry. SWAFS Journal. 2013; 34: 7-30.
- 7. Lum B. Distinguishing Isomers of 3-Fluoromethcathinone (3-FMC) from 4-Fluoromethcathinone (4-FMC) using Heptafluorobutyric anhydride (HFBA) chemical derivatization. SWAFS Journal. 2012; 33: 6-17.
- 8. SWGDRUG. *Monographs: 2-Fluoromethcathinone.* Retrieved from http://www.swgdrug.org/monographs.htm. 2013.





the TIC of 3-FMC.







• • • 50



1.997 minutes in FluoroPFPAMix.









Mix.





Fig. B19: Mass spectrum for Methody PFPA at 7.501 minutes in Methoxy PFPA Mix.



Appendix C: Additional Methylmethcathinone Data



Fig. C13: Mass spectrum of 3-MMC-HFBA at 3.313 minutes in Methyl HFBA Mix.



Mix.



Fig. D6: Mass spectrum of 10.289 peak from the TIC of 2,3-MDMC-TFAA.



Fig. D9: Mass spectrum of 10.056 peak from the TIC of 2,3-MDMC-HFBA.

Fig. D12: Mass spectrum of Methylone-HFBA at 8.674 minutes in Methylenedioxy HFBA Mix.





the TIC of 2~MPPP.

the TIC of 4~MPPP.

