Development and Internal Validation of a Chelex[®] DNA Extraction Protocol for Reference Oral Swabs Choon Sung Kambara, B.S.¹, Rebecca Boissaye, M.S.F.S.², Joshua Stewart, M.S.F.S.¹, and Pamela Staton, Ph.D.¹ ¹ Marshall University Forensic Science Center, 1401 Forensic Science Drive, Huntington, WV 25701 ²Boston Police Department Crime Laboratory, 1 Schroeder Plaza, Boston, MA 02120



Abstract

Forensic DNA laboratories have limited budgets, analysts and time with an abundance of samples to be examined. Therefore, it is necessary to implement procedures in a lab that maximize efficiency while minimizing cost. Additionally, any procedure used in a lab must be validated following SWGDAM's Validation Guidelines to show it is a reliable and robust method and to determine any limitations. The purpose of this research was to devise and validate such a procedure for the Boston Police Department Crime Laboratory DNA Section. Specifically, a BioRad Chelex[®] resin based extraction procedure for reference oral swabs.

Introduction

This project had three stages: method development, validation and addressing any additional questions that arose during the course of the project.

Development Studies

–Swab sampling type

-Room temperature incubation length

–56°C incubation length

–Boiling method

Validation Studies

-Concordance

-Contamination risk assessment

–Ability of Chelex[®] to extract DNA from potentially compromised swabs – artificially aged swabs

- Original and revised protocol
- –Stability of Chelex[®] extract

Studies to Address Additional Questions

–Stability of 5% Chelex[®] Solution

- –Amplification target assessment
- –Post-Extraction interference studies
 - Biological sample type Blood vs. Saliva
 - Extraction method EZ1[®] vs. Chelex[®]
 - Sample substrate type Liquid vs. Stained Swab

Materials and Methods

Kits, Instruments and Software:

- -BioRad BT Chelex[®] 100 Resin
- –Qiagen EZ1[®] Advanced
- –EZ1[®] DNA Investigator Kit
- –Applied Biosystems[®] Quantifiler[®] Duo Quantification Kit
- –Applied Biosystems[®] 7500 Real-Time PCR system
- –AmpFLSTR[®] Identifiler Plus[®] Amplification Kit
- –Applied Biosystems[®] 9700 Thermal Cycler
- -Applied Biosystems[®] 3130xl Genetic Analyzer
- –GeneMapper[®] ID v3.2.1

Sample Materials:

–Previously collected oral swabs refrigerated for 0-5 years

- -Freshly collected oral swabs from known donors
- –Saliva samples from known donors
- -Control rbs fabrics of known donor

-Liquid blood control samples from serology section

Method Deve	lopment Res	ults
Parameter Evaluated	Original Protocol	Developed Protocol
Swab Sample Type	Undefined for Swab	Tip of Swab Cutting
Room Temperature Incubation Length	15-30 min	15 min
56°C Incubation Length	15-30 min	15 min
Boiling Method	Boiling Water Bath	100°C Dry Bath
Method Vali	dation Resu	lts
Concordance –All samples of known profile	were found to be	concordant.
–All reagent blanks met BPD's uncontaminated reagent blan	e nt requirements for ks.	
Stability Study: Substrate Var	iability – Revised	Procedure
0.5 ng Day 0 Day 1	5 ■ Day 34 ■ D	Pay 50
Full Profile Pull Up Heteroz Imb	zygote PHR Stutter Dalance	Additional Artifacts
Percent of	Profiles with:	
Figure 1 compares characterist from swabs that were artificial in a 37°C incubator and then ex Chelex [®] procedure.	tics of DNA profile ly aged for 0, 15, 3 xtracted with the	s obtained 84 and 50 days revised
Stability Study: Substrate Variation - Similar results to Revised Proc	ability – Original I cedure Study.	Procedure
Extract Stability Study		
Figure 2: Comparison of multi amplified wi	ple amplifications of Dation of D	ay 0 extract
■ Amp 1 ■ Amp 10 10 10 10	o 2 🔳 Amp 3 🗖 Amp	o 4
rui Frome Fui Op Heter Ir	mbalance	Additional Altilacts
Percent o	f Profiles with:	
Figure 2 compares character refrigeration of Day 0 (ori approximately 0, 15, 34 and 50)	eristics of DNA ginal procedure) days.	profiles after extracts for

Additional Studies Results

% Chelex[®] Solution Stability Study



olution up to 272 day after preparation.

Amplification Target Assessment Study



igure 4 compares the quality of the DNA profiles obtained rom oral swabs when extracted with Chelex[®] and amplified vith either 0.25 ng or 0.5 ng of DNA.

Post-Extraction Interference Studies

lypothesis: Source of disconnect between quantification alues and amplification results is due to underestimation of uantification results due one or a combination of:

- sample type
- substrate type
- extraction method.

Sample Type – blood vs. saliva

Relative underestimation of blood

Extraction Method – Chelex[®] vs. EZ1[®]

Slight indication of relative underestimation of Chelex[®]

ubstrate Type – liquid vs. stained swabs

Relative underestimation of stained swabs

Cost Analysis Assessment

Cost of Kit/Bottle Number of Samples per Kit/Bottle Cost per Sample

Qiagen DNA Investigator Kit \$414.72 48 \$8.64

Chelex[®] 100 Resin (100 g Bottle) \$397.10 11765

\$0.03

A protocol was developed based on BPD's Chelex[®] DNA Extraction from Whole Blood/Bloodstains/Saliva Protocol and previous studies performed by the BPD crime lab DNA section. The changes made in the development of the extraction protocol included use of a tip of swab cutting, a 15 minute room temperature incubation, a 15 minute 56°C incubation and use of a dry bath set at 100°C.

This protocol was evaluated to determine whether it was a reliable method and to determine its limitations.

-Sensitivity of this protocol is sufficient to extract amplifiable quantities of DNA from oral swabs in sufficient quality to be used for DNA analysis and provide interpretable, concordant DNA profiles.

-Capable of extracting DNA from artificially-aged swabs.

-Extract is stable when stored at 2-8°C for at least 50 days.

Additional studies were performed to answer questions that arose during the project.

-An intermediate amplification target amount, perhaps 0.35 ng, should be evaluated.

-5% Chelex[®] solution should be made more than once a year with a pH monitoring system set in place.

–Possible relative underestimation of quantification values is associated with stained swabs (compared to liquid samples) and blood based samples (over saliva samples).

-Slight indication that Chelex[®] extracted samples have underestimation in quantification values compared to EZ1 extracted samples.

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Conclusions

-Low risk of contamination

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